

Ning-Juan Fan,<sup>a</sup> Dong-Hong Li,<sup>a</sup>  
Xiao-Ming Jiang,<sup>b</sup> Yong-Bin  
Zhang,<sup>a</sup> Wei Guo<sup>a\*</sup> and  
Chi-Zhong Xia<sup>a</sup>

<sup>a</sup>School of Chemistry and Chemical Engineering, University of Shanxi, Taiyuan, Shanxi 030006, People's Republic of China, and <sup>b</sup>Institute of Molecular Science, Chemical Biology and Molecular Engineering Laboratory of the Education Ministry, Shanxi University, Taiyuan, Shanxi 030006, People's Republic of China

Correspondence e-mail: guow@sxu.edu.cn

#### Key indicators

Single-crystal X-ray study  
 $T = 183\text{ K}$   
Mean  $\sigma(\text{C}-\text{C}) = 0.002\text{ \AA}$   
 $R$  factor = 0.043  
 $wR$  factor = 0.111  
Data-to-parameter ratio = 13.1

For details of how these key indicators were automatically derived from the article, see <http://journals.iucr.org/e>.

## Ethyl 2-acetyl-3-[2-(1*H*-indol-3-yl)ethyl-amino]propenoate

The title compound,  $\text{C}_{17}\text{H}_{20}\text{N}_2\text{O}_3$ , was synthesized by the substituted one-carbon unit transfer reaction of tryptamine with an imidazolidine derivative. There are intramolecular and intermolecular  $\text{N}-\text{H}\cdots\text{O}$  hydrogen bonds in the crystal structure.

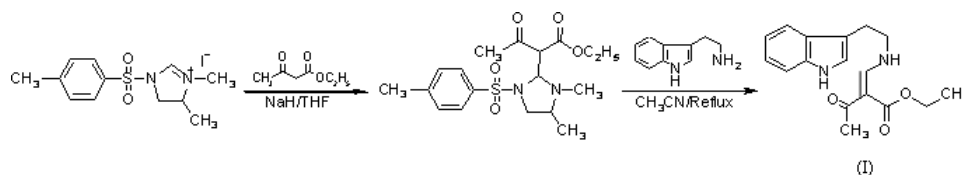
Received 7 December 2004

Accepted 13 January 2005

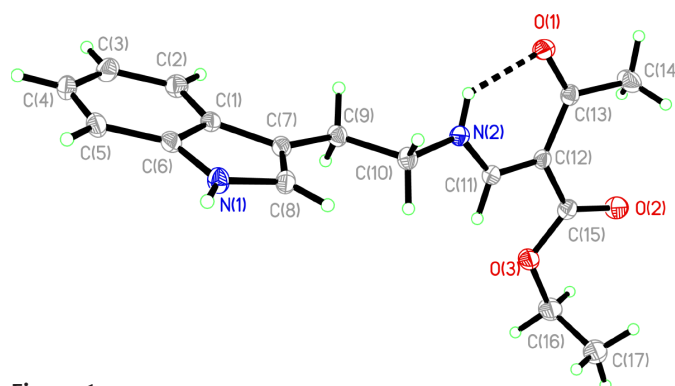
Online 22 January 2005

#### Comment

Secondary enamines have attracted a great deal of attention in recent years because of their range of applications (Duthaler, 2003; Stanovnik & Svete, 2004; Elassar & El-Khair, 2003). Moreover, functionalized secondary enamine derivatives may enable chemical and biological studies on these derivatives, which will be used in pharmaceutical research. We were therefore interested in the synthesis of secondary enamine derivatives during our investigation of tetrahydrofolate coenzyme models (Li *et al.*, 2004). Tetrahydrofolate is involved in the biological transfer of a carbon unit at different oxidation levels (Blakley, 1969), which can transfer  $-\text{C}=\text{C}-$  or  $=\text{CH}-$  groups *via* mechanisms analogous to those operative in biochemical processes (Bieraugel *et al.*, 1983; Pandit & Bieraugel, 1979).

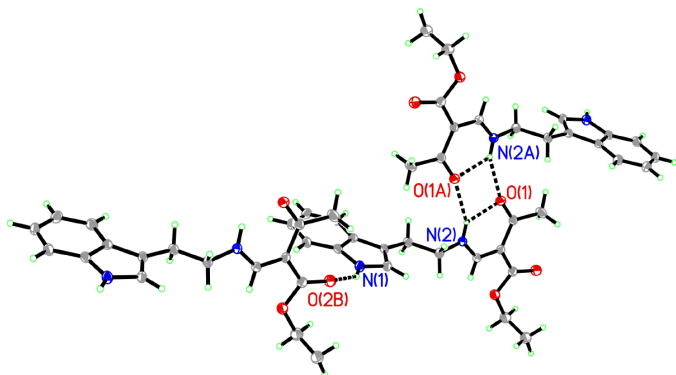


The title compound, (I), was obtained by the substituted one-carbon unit transfer reaction of tryptamine with the imidazolidine derivative which was produced by the addition reaction of 1-tosyl-3,4-dimethylimidazolium iodide with a carbanion of acetylacetate. This might be regarded as a



**Figure 1**

The molecular structure of (I), showing 50% probability ellipsoids. The dashed line indicates a hydrogen bond.



**Figure 2**

The three-centered hydrogen bonds between two O atoms of two carbonyl groups and atom N2. [Symmetry codes: (A)  $-x, 1-y, 1-z$ ; (B)  $x, y-1, z+1$ .]

substituted one-carbon unit transfer reaction which mimics the function of tetrahydrofolate coenzymes.

The molecular conformation of (I) is illustrated in Fig. 1, and some features of the geometry are listed in Table 1. A number of intra- and intermolecular hydrogen bonds (Table 2) stabilize the crystal structure. Three-centered hydrogen bonds are formed between two O atoms of two carbonyl groups and atom N2 (Fig. 2).

## Experimental

Sodium hydride (1.8 mmol) was added to a solution of acetyl acetate (1.5 mmol) in dry tetrahydrofuran (10 ml), which was cooled in an ice–water bath. The reaction mixture was stirred for 30 min, then 1-tosyl-3,4-dimethylimidazolinium iodide (1 mmol) was added and the mixture was allowed to warm to room temperature and stirred continuously for 3 h, then quenched with water. The solution was extracted with dichloromethane, dried over anhydrous sodium sulfate and concentrated. The residue was purified by column chromatography to afford a yellow oil, (II). The imidazolidine derivative, (II) (1 mmol), and tryptamine (1 mmol) in anhydrous acetonitrile (10 ml) were refluxed, the solution concentrated and the residue purified by column chromatography to afford compound (I) (yield 85%). Crystals suitable for X-ray analysis were obtained by slow evaporation of an ethanol solution.

### Crystal data

$C_{17}H_{20}N_2O_3$   
 $M_r = 300.35$   
 Triclinic,  $P\bar{1}$   
 $a = 8.4342$  (18) Å  
 $b = 9.391$  (2) Å  
 $c = 10.360$  (2) Å  
 $\alpha = 79.941$  (3)°  
 $\beta = 71.597$  (3)°  
 $\gamma = 89.332$  (3)°  
 $V = 765.8$  (3) Å<sup>3</sup>

$Z = 2$   
 $D_x = 1.303$  Mg m<sup>-3</sup>  
 Mo  $K\alpha$  radiation  
 Cell parameters from 1672 reflections  
 $\theta = 2.5$ – $26.9$ °  
 $\mu = 0.09$  mm<sup>-1</sup>  
 $T = 183$  (2) K  
 Block, colorless  
 $0.40 \times 0.30 \times 0.30$  mm

### Data collection

Bruker SMART 1K CCD area-detector diffractometer  
 $\varphi$  and  $\omega$  scans  
 Absorption correction: multi-scan (SADABS; Sheldrick, 1996)  
 $T_{\min} = 0.494$ ,  $T_{\max} = 0.973$   
 3165 measured reflections

2636 independent reflections  
 2154 reflections with  $I > 2\sigma(I)$   
 $R_{\text{int}} = 0.015$   
 $\theta_{\text{max}} = 25.0$ °  
 $h = -10 \rightarrow 8$   
 $k = -10 \rightarrow 11$   
 $l = -12 \rightarrow 12$

### Refinement

Refinement on  $F^2$   
 $R[F^2 > 2\sigma(F^2)] = 0.043$   
 $wR(F^2) = 0.111$   
 $S = 1.01$   
 2636 reflections  
 201 parameters

H-atom parameters constrained  
 $w = 1/[\sigma^2(F_o^2) + (0.0645P)^2]$   
 where  $P = (F_o^2 + 2F_c^2)/3$   
 $(\Delta/\sigma)_{\text{max}} < 0.001$   
 $\Delta\rho_{\text{max}} = 0.20$  e Å<sup>-3</sup>  
 $\Delta\rho_{\text{min}} = -0.18$  e Å<sup>-3</sup>

**Table 1**

Selected geometric parameters (Å, °).

O1–C13	1.2358 (19)	N1–C6	1.368 (2)
O2–C15	1.2105 (19)	N1–C8	1.369 (2)
O3–C15	1.347 (2)	N2–C11	1.2998 (19)
O3–C16	1.448 (2)	N2–C10	1.4556 (19)
C1–C7–C9–C10	171.15 (15)	C7–C9–C10–N2	–168.51 (13)
C11–N2–C10–C9	–110.86 (15)	C10–N2–C11–C12	–172.13 (15)

**Table 2**

Hydrogen-bonding geometry (Å, °).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
N2–H2 $\cdots$ O1	0.88	2.02	2.6518 (17)	128
N2–H2 $\cdots$ O1 <sup>i</sup>	0.88	2.32	3.0393 (18)	139
N1–H1 $\cdots$ O2 <sup>ii</sup>	0.88	2.07	2.8572 (17)	149

Symmetry codes: (i)  $-x, 1-y, 1-z$ ; (ii)  $x, y-1, 1+z$ .

H atoms attached to C and N atoms were placed in geometrically idealized positions, and constrained to ride on their parent atoms, with C–H = 0.93–0.96 Å and N–H = 0.88 Å, and with  $U_{\text{iso}}(\text{H}) = 1.2U_{\text{eq}}(\text{parent atom})$ .

Data collection: SMART (Bruker, 2000); cell refinement: SAINT (Bruker, 2000); data reduction: SAINT; program(s) used to solve structure: SHELXS97 (Sheldrick, 1997); program(s) used to refine structure: SHELXL97 (Sheldrick, 1997); molecular graphics: SHELXTL/PC (Sheldrick, 1999); software used to prepare material for publication: SHELXTL/PC.

This work was supported by the National Natural Science Foundation of China (No. 20272034, to CZX), and the Natural Science Foundation of Shanxi Province, China (Nos. 20041007 and 20041006).

## References

- Bieraugel, H., Plemp, R., Hiemstra, H. C. & Pandit, U. K. (1983). *Tetrahedron*, **39**, 3971–3979.
- Blakley, R. L. (1969). *The Biochemistry of Folic Acid and Related Pteridines*. New York: American Elsevier.
- Bruker (2000). SMART (Version 5.0) and SAINT (Version 6.02). Bruker AXS Inc., Madison, Wisconsin, USA.
- Duthaler, R. O. (2003). *Angew. Chem. Int. Ed.* **42**, 975–978.
- Elassar, A. Z. A. & El-Khair, A. A. (2003). *Tetrahedron*, **59**, 8463–8480.
- Li, D., Hao, J., Guo, W. & Xia, C. (2004). *Heterocycles*, **63**, 2515–2522.
- Pandit, U. K. & Bieraugel, H. (1979). *J. Chem. Soc. Chem. Commun.* pp. 117–119.
- Sheldrick, G. M. (1996). SADABS. University of Göttingen, Germany.
- Sheldrick, G. M. (1997). SHELXL97 and SHELXS97. University of Göttingen, Germany.
- Sheldrick, G. M. (1999). SHELXTL/PC. Version 6.10. Bruker AXS Inc., Madison, Wisconsin, USA.
- Stanovnik, B. & Svete, J. (2004). *Chem. Rev.* **104**, 2433–2480.